## SYNTHESIS OF INOSINATE TRIMER I2'p5'I2'p5'I AND TETRAMER I2'p5'I2'p5'I2'p5'I

RAMAMURTHY CHARUBALA AND WOLFGANG PFLEIDERER Fakultät für Chemie, Universität Konstanz, Postfach 5560, D-7750 Konstanz/West Germany

The trimeric and tetrameric inosinates with (2'-5')-linkages were synthesized via the phosphotriester approach using the p-nitrophenylethyl group for phosphate protection.

The exciting reports [1-5] on the unusual structure of the oligonucleotide-5'-O-triphosphoryladenylyl(2'- 5')adenylyl(2'- 5')adenosine (ppA2'p5'A2'p5'A) and its biological activity as a strong inhibitor of cell free protein synthesis forced various research groups all over the world to synthesize this low-molecular-weight oligonucleotide [6-11] as well as its core A2'p5'-A2'p5'A [12-17] by a purely chemical approach using in general the phosphotriester method but varying the protecting group combinations at the various positions of the carbohydrate moiety and the aglycone. The corresponding 3'-deoxyadenylate trimer dA2'p5'A2'p5'A has recently been synthesized in our laboratory [18] and was found [19] to act as a substitute for human fibroblast interferon in those cells that are permeable to core nucleotides. It is further of particular interest to know that the core cordycepin trimer represents so far the most valuable candidate of the new class of (2'- 5')oligonucleotides owing to its antiviral effect coupled with its extended metabolic stability without toxicity to cells.

The availability of various oligomers with 2'- 5' internucleotidic linkages would facilitate further studies of their biological action especially if such structural analog as the deaminated counterparts are taken into consideration. First results on these lines have been obtained recently [20] during the polymerization of inosine-5'-phosphorimidazolide in presence of Pb<sup>++</sup> ions but with low yields of the trimer and tetramer. In this paper we wish to describe therefore a direct chemical synthesis of the 2'-5'-inosinate trimer and tetramer using the phosphotriester approach and the p-nitrophenylethyl group for phosphate protection [21,22] as the key features.

Inosine  $(\underline{1})$  was first blocked in 5'-position by the monomethoxytrityl group

4789



to give  $\underline{2}$  in 80 % yield. Its treatment with t-butyldimethylsilylchloride and imidazole in pyridine led to a mixture of 40 % of the 2'-O-( $\underline{5}$ ) and 3'-O-tbutyldimethylsilyl derivative ( $\underline{6}$ ) each and minor amounts (5-8 %) of the 2',3'disilylated analog ( $\underline{7}$ ). Phosphorylation of  $\underline{6}$  was carried out using 2,5-dichlorophenyl-phosphoroditriazolide in pyridine followed by p-nitrophenylethanol to give the corresponding 2'-phosphotriester  $\underline{10}$  in 95 % yield after chromatography on a silica gel column.  $\underline{10}$  functioned as a versatile synthon since oximate cleavage [23] deblocked quantitatively the 2,5-dichlorophenyl group to form the phosphodiester  $\underline{11}$ , whereas deprotection of the monomethoxytrityl group by 2 % p-toluene sulfonic acid in methylenechloride/methanol (7/3) gave 3'-O-t-butyldimethylsilylinosine-2'-(2,5-dichlorphenyl, p-nitrophenylethyl)phosphate ( $\underline{9}$ ) in 92 % yield. 2',3'-Di-O-t-butyldimethylsilyl-( $\underline{8}$ ) and 2',3'di-O-benzoylinosine ( $\underline{4}$ ) were prepared from  $\underline{2}$  by silylation and benzoylation respectively followed by detritylation.

The synthesis of the inosinate trimer 21 started from 8 and 11, which have first been coupled by quinoline-8-sulfonyl chloride (QSCl) and 3-nitro-1,2,4triazole (NT) (1/3) in absol. pyridine at room temp. for 16 h to give the dimer 16 in 79 % isolated yield. Cleavage of the monomethoxytrityl group yielded 88 % of 17, which was then condensed in a similar manner with 11 to the fully protected inosylyl(2'- 5')inosylyl(2'- 5')inosine (19) in 88 % yield.

The preparation of the inosinate tetramer  $\underline{20}$  was achieved by block-condensation of the two dimers  $\underline{13}$  and  $\underline{15}$ .  $\underline{13}$  was obtained from the condensation reaction of the phosphodiester  $\underline{11}$  with  $\underline{4}$  first to  $\underline{12}$  in 76 % and subsequent detritylation, whereas  $\underline{15}$  resulted from the reaction between  $\underline{9}$  and  $\underline{11}$  to  $\underline{14}$  in 71 % yield and followed by oximate cleavage of the 2,5-dichlorophenyl group. The coupling reaction was performed under standard conditions applying QSC1 and NT in pyridine to form the fully protected tetrameric inosine  $\underline{18}$  in 81 % yield.

Both oligomers  $\underline{18}$  and  $\underline{19}$  were then deblocked using first 0.5 M DBU in absol. pyridine at room temp. for 6-8 h to remove the p-nitrophenylethyl group by elimination and second 0.5 M tetrabutylammonium fluoride for cleavage of the silyl groups. In  $\underline{18}$  conc. ammonia was then applied for benzoyl deprotection, whereas the detritylation was always performed in the last step by 80 % acetic acid in 6-8 h. The resulting products were put on a DEAE-Sephadex A-25 column and were eluted with a linear gradient of 0.001M - 0.6M triethylammonium bicarbonate buffer pH 7.5. The isolation of pure inosinate trimer  $\underline{21}$  and tetramer  $\underline{20}$  respectively resulted from the main fractions by several coevaporations with water and final lyophylisation to stable amorphous powders in 91 % yield each. - The purity of all products has been checked by chromatographical means and the various structures have been proven by UV and NMR spectra as well as elementary analyses.

## REFERENCES

1)	A.G. Hovanessian, R.E. Brown and I.M. Kerr, <u>Nature (London)</u> <u>268</u> , 537 (1977).
2)	I.M. Kerr, R.E. Brown and A.G. Hovanessian, <u>Nature (London)</u> <u>268</u> , 540 (1977).
3)	I.M. Kerr and R.E. Brown, Proc.Natl.Acad.Sci. USA 75, 256 (1978).
4)	E.M. Martin, J.M. Birdsall, R.E. Brown and I.M. Kerr, <u>Eur.J.Biochem</u> . <u>95</u> , 295 (1979).
5)	C.M. Vaquero and M.J. Clemens, Eur.J.Biochem. 98, 245 (1978).
6)	M. Ikehara, K. Oshie and E. Ohtsuka, <u>Tetrahedron Lett.</u> 20, 3677 (1979).
7)	H. Sawai, T. Shibata and M. Ohno, <u>Tetrahedron Lett.</u> 20, 4573 (1979).
8)	J.F.M. Rooij, G.W. Hazeleger, P.H. van Deursen, J. Serdjin and J.H. van Boom, <u>Recl.Trav.Chim. Pays-Bas</u> <u>98</u> , 537 (1979).
9)	S.S. Jones and C.B. Reese, <u>J.Am.Chem.Soc.</u> <u>101</u> , 7399 (1979).
10)	J. Imai and P.F. Torrence, <u>J.Org.Chem.</u> <u>46</u> , 4062 (1981).
11)	E. Ohtsuka, A. Yamane and M. Ikehara, <u>Chem.Pharm.Bull.</u> <u>30</u> , 376 (1982).
12)	K.K. Ogilvie and N.Y. Theriault, <u>Tetrahedron Lett</u> . 20, 2111 (1979).
13)	A.K. Markham, R.A. Porter, M.J. Gait, R.C. Sheppard and I.M. Kerr, <u>Nucleic Acids Res</u> . <u>6</u> , 2569 (1979).
14)	J. Engels and U. Krahmer, <u>Angew.Chem.</u> <u>91</u> , 1007 (1979).
15)	R. Charubala and W. Pfleiderer, <u>Tetrahedron Lett.</u> 21, 1933 (1980).
16)	C. Gidelli, M. Kwiatkowski, B. Öberg and J.B. Chattopadhyaya, <u>Tetra-</u> <u>hedron Lett</u> . <u>22</u> , 1741 (1981).
17)	R. Charubala, E. Uhlmann and W. Pfleiderer, <u>Liebigs Ann.Chem.</u> <u>1981</u> , 2392.
18)	R. Charubala and W. Pfleiderer, <u>Tetrahedron Lett</u> . <u>21</u> , 4077 (1980).
19)	P.W. Doetsch, R.J. Suhadolnik, Y. Sawada, J.D. Mosca, M.B. Flick, N.L. Reichenbach, A.Q. Dang, J.M. Wu, R. Charubala, W. Pfleiderer and E.E. Henderson, <u>Proc.Natl.Acad.Sci</u> . <u>USA</u> , <u>78</u> , 6699 (1981).
20)	H. Sawai and M. Ohno, <u>Bull.Chem.Soc.Jpn.</u> 54, 2759 (1981).
21)	E. Uhlmann and W. Pfleiderer, <u>Tetrahedron Lett</u> . <u>21</u> , 1181 (1980).
22)	E. Uhlmann and W. Pfleiderer, <u>Helv.Chim. Acta</u> <u>64</u> , 1688 (1981).
23)	C.B. Reese and L. Zard, <u>Nucleic Acids Res.</u> 9, 4611 (1981).
	(Received in Germany 3 August 1982)